

# Efficacy of certain fungicide alternatives for controlling sugar beet powdery mildew

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#### Abstract

In order to study the importance of selection of suitable sugar beet cultivars for growing in Upper Egypt, ten sugar beet cultivars were tested for their susceptibility to Erysiphe betae, the causal fungus of powdery mildew Among them, Sirona (453 AUPMPC; the Area Under Powdery Mildew Progress Curve) was significantly the most resistant cultivar to powdery mildew disease while, FD.0807 (1484 AUPMPC) was the most susceptible one. Field experiment was conducted to evaluate five plant extracts, three chemical antioxidants and four microelement compounds for their efficacy in controlling E. betae. Results confirmed that all tested treatments caused significant reduction in AUPMPC values when sugar beet plants were sprayed with these tested compounds and the increasing of the concentration increased resistance of sugar beet plants against powdery mildew disease. Among the tested plant extracts, the highest protection (69.9% & 66.9%) on both cultivars Sirona and FD.0807 respectively was achieved by 30% of basil extract followed by 30% of garlic extract (53% & 60.4%) while, the least protection (33.9% & 52.8%) was obtained by 10% of black cumin seed extract. Concerning the tested inducers, the highest protection (56.7% & 70.6%) was achieved by 300 ppm of salicylic acid followed by 200 ppm of salicylic acid (51% & 63.7%), while, 100 ppm of ascorbic acid gave the least protection (2.2% & 34.1%). On the other hand, 40 ppm of cupric sulfate achieved the best percentage of protection (72% &78.7%) whereas, the least protection (8%) was obtained by 10 ppm of zinc sulfate on Sirona cultivar and the least protection (43.6%) was obtained by 10 ppm of magnesium sulfate on FD.0807 cultivar. The best comparative treatment was the tested fungicide Bellis® 38% WG (89.5% &90% protection). Usage of plant extracts, antioxidants and micro elements are well recommended as fungicide alternatives for controlling the disease in parallel with their safe influence on human health due to reduction of the accumulated chemical hazards in the plant tissues.

Keywords: sugar beet, powdery mildew, Erysiphe betae, plant extracts, micronutrients.



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### Introduction

Powdery mildew of sugar beet caused by Erysiphe betae (Vanha) Weltzien is a common disease in many countries (Hills et al., 1980). In Egypt, environmental conditions are favorable for the fungus to spread rapidly especially in the late sowings after September (El-Fahhar, 2008).Disease spread occurs mostly by conidial infections, which dispersed by wind (Kontradowitz & Verreet, 2010). The infection of sugar beet by powdery mildew is economically significant for growers worldwide and can cause sugar yield losses up to 30% 2002). Utilizing (Francis. disease resistance is an important strategy for controlling plant pathogens because it may be relatively easy to use, costeffective and environmentally more acceptable than reliance on pesticides (Hogenboom, 1993). Moderate resistance to the disease is obtainable in commercial varieties appearing in "slow-mildewing" phenotypes (Kontradowitz & Verreet, 2010). Many chemical constituents extracted from different plant genera were tested in the past years and found to be successful in controlling powdery mildew fungi on several crops (Moharam & Ali, 2012; El-Fahhar et al., 2009). The activation of a wide range of disease resistance mechanisms and the production of a wide range of defense compounds; it is race non-specific and is often effective against a broad spectrum of pathogenic agents (Ismail et al., 2012; Galal & Abdou, 1996; Kuc, 1995; Elad, 1992). The objective of this work was to evaluate some sugar beet cultivars for their susceptibility to powdery mildew disease and to evaluate some chemical and plant extracts for controlling the disease under field conditions.

### Materials and methods

Reaction of ten sugar beet cultivars to powdery mildew: Field experiment was carried out at the experimental field of the Faculty of Agriculture, Al-Azhar University, Assiut governorate, Egypt, 2013/2014 during and 2014/2015 growing seasons. Seeds of ten sugar beet cultivars (Belatos, Capel, Oscarpoly, FD.0807, Pleno. Glorius. Sirona. Hilospoly, Samba and Athospoly) which were kindly gifted from Agricultural Research Station, Sakha, Kafr El-Sheikh were sown in plots consisted of ten rows (9-meters long, interspace between plant and another 20 cm and 3 seeds/hole were sown) in a completely randomized design with three replicates of each cultivar. Sugar beet plants were fertilized and irrigated as recommended by Egyptian Ministry of Agriculture. Plants were thinned to one plant/hole and left for natural infection. Disease severity was estimated three times, when plants were closing the rows (120 days after sowing) then estimated twice again (140 and 160 days after sowing) to calculate the Area Under Powdery Mildew Progress Curve (AUPMPC) values.

Powdery mildew assessment: Evaluation of disease severity was accomplished by examining both sides of leaves and rating disease intensity as the extent of leaf area covered by the fungus mycelium on a scale of 0 to 4 according to the scale by Whitney et al., (1983). Scale ranged from 0-4, categories whereas 0 =no mildew colonies observed, 1=1-25%, 2=26-50%, 3=51-75% and 4=76-100% of matured leaf area covered by mildew.

Area Under Powdery Mildew Progress Curve (AUPMPC) was calculated for the assessment period using the following equation adopted by Chiha *et al.* (1997).

AUPMPC = D 
$$(1/2 (Y_1 + Y_k) + (Y_2 + Y_3 + \dots Y_{k-1}))$$

Where:  $D = Time interval; Y_1 = First$ disease score;  $Y_k = Last$  disease score;  $Y_2$ and  $Y_3 = Intermediate$  disease score.

Evaluation of some plant extracts, antioxidants and micronutrients for controlling sugar beet powdery mildew: Field experiment was carried out at the experimental field of the Faculty of Agriculture, Al-Azhar Univ., Governorate, Assiut Egypt, during 2014/2015 and 2015/2016 growing seasons. Sirona and FD.0807 sugar beet cultivars were selected for the experiment which exhibited resistant and susceptible reaction, respectively to powdery mildew disease based on their interactions in the previous experiment in the reaction of sugar beet cultivar. Field plots consisted of two rows (9-meters long and interspace between plant and another 20 cm) and arranged in a split plot design with three replicates per treatment. One plot was specified for each tested compound and one plot was left as a check for all treatments. Agricultural practices were applied as recommended by Ministry of Agriculture. Plants were thinned to one plant / hole and left for natural infection. Large area around the plots was left without treatment to avoid any contamination by any treated chemicals from neighboring fields (Gado, 2013).

**Preparation of the tested compounds:** Aqueous plant extracts were prepared from all parts (leaves, seeds and /or cloves) of five plants *i.e.*, basil, camphor, garlic, black cumin and mint as following:

Tested plants	The used part
Basil (Ocimumbasilicum)	Leaves
Camphor (Eucalyptus globulus)	Leaves
Garlic (Allium sativum)	Cloves
Black cumin (Nigella sativa)	Seeds
Mint (Mentha piperita)	Leaves

The plant parts were washed several times with sterilized distilled water, cut into small pieces, then 100 g of each sample were macerated in 100 ml sterilized distilled water by using mortar, after that, resulting extract was squeezed twice through four layers of cheese cloth as described by Hassan (2006) with slight modification. Filtrates of aqueous plant crude extracts were kept in dark bottles in refrigerator until used. Each aqueous plant extract was used as foliar spraying at the concentrations of 10, 20 and 30 % attributed to the crude extract. Sugar beet plants were sprayed after 105, 125, 145, 165 and 185 days from sowing date.

The efficacy of three chemical inducers *i.e.* ascorbic acid, salicylic acid and citric acid were tested to study their effect against powdery mildew disease of sugar beet plants (Sirona cv. and F.D.0807 cv.). Each chemical inducer was used as foliar spraying at the concentrations of 100, 200 and 300 ppm (Ismail *et al.*, 2012). Sugar beet plants were sprayed after 105, 125, 145, 165 and 185 days from sowing date.

To assess the efficiency of microelements, as foliar sprays on the control of powdery mildew disease on

sugar beet plants (Sirona cv. and F.D.0807 cv.), solutions of cupric sulfate, ferrous sulfate, magnesium sulfate, zinc sulfate and mixture of all of them at the concentrations of 10, 20 and 40 ppm (Radwan, 2017) were used as a foliar spray after 105, 125, 145, 165 and 185 days from sowing date.

The used fungicide was commercial formulation of 25.2% w/w boscalid and 12.8% w/w pyraclostrobin (Bellis® 38% WG). Fungicide was applied in the dosage (0.5 gm / liter) as cited in its user manual sheet which is recommended by the manufacturer (BASF<sup>TM</sup>) as described by Moustafa et al. (1990).

Time application: of Treatment applications were started 105 days after sowing (the first sign of the disease has appeared). Plants were sprayed five times during each season with 20 days intervals. Tap water was used for spraying untreated plants (control). Disease severity was determined (five times) in order to evaluate treatments after ten days from each spraying of tested compounds. Solutions of each tested compounds were applied using a hand sprayer, at a volume of 2 Liters of tap water per plot (until run off). Thirty plants were used for each treatment. Plants without spraying served as control. AUPMPC values were calculated as described before.

**Statistical analysis:** Analysis of variance of the data was carried out on the mean values of the tested treatments according to the procedures described by Gomez and Gomez (1984). The least significant difference (L.S.D) at 5% probability was used for testing the significance of the differences among the mean values of the tested treatments for each character.

## Results

Reaction of sugar beet cultivars for powdery mildew infection: Most of tested sugar beet cultivars significantly responded with varied degrees to powdery mildew infection (Table 1). Plants of line FD.0807 were most susceptible followed by Athospoly cv., Hilospoly cv., Oscarpoly cv., Belatos cv. and Pleno cv. respectively. Capel cv., Glorius cv. and Samba cv. showed intermediate reaction to powdery mildew while, Sirona cv. showed the lowest AUPMPC value. In that trend of reactions to the disease, all cultivars took place in the same orders during both growing seasons tested.

Table 1: Mean of area under powdery mildew progress curve (AUPMPC) values for ten sugar beet cultivars during 2013/2014 and 2014/2015 growing seasons under natural infection.

Cultivars	AUPMPC						
Cultivars	2013/2014	2014/2015	Mean				
Athospoly	1315	1312	1313				
Belatos	917	939	928				
Capel	798	757	777				
FD.0807	1467	1502	1484				
Glorius	703	718	710				
Hilospoly	1090	1125	1107				
Oscarpoly	981	942	961				
Pleno	895	861	878				
Samba	673	676	674				
Sirona	443	462	453				
LSD at 0.05 %	36.7	55.7					

Efficacy of plant extracts to natural sugar beet powdery mildew: Data in (Table 2) showed that the five plant extracts at the three tested concentrations 10, 20 and 30% reduced the AUPMPC significantly during both 2014/2015 and 2015/2016 growing seasons as compared

with the control but each to a different extent. The concentration of 30% was more effective in reducing AUPMPC than the concentrations of 20% and 10% respectively in each particular plant extract. Repeated sprays of basil extract at the concentration of 30% significantly achieved the lowest AUPMPC on both sugar beet cultivars Sirona and FD.0807, followed by garlic extract. Moderate effects were achieved by camphor and mint extracts while, black cumin seed extract came in the last order. The best treatment (Basil 30%) was higher in AUPMPC than the tested fungicide Bellis® 38% WG.

Table 2: Mean of AUPMPC values to sugar beet (Sirona & FD.0807 cultivars) as affected by foliar spray with plant extracts under field conditions during 2014/2015 and 2015/2016 growing seasons.

		AUPMPC								
Treatment	Conc.		Sirona	cv.		FD.0807 L.				
Treatment	(%)	2014/2015	2015/2016	Mean	Protection (%)	2014/2015	2015/2016	Mean	Protection (%)	
	10	508	496	$502 \pm 1$	28	842	851	847 ±1	48.7	
Basil	20	401	406	$403 \pm 1$	42.2	703	724	$713 \pm 1$	56.8	
	30	218	203	$210 \pm 1$	69.9	553	538	$546 \pm 1$	66.9	
	10	700	657	$678 \pm 1$	2.8	1212	1165	$1188 \pm 1$	28.1	
Black cumin	20	541	515	$528 \pm 1$	24.3	927	929	$928 \pm 1$	43.8	
	30	474	448	$461 \pm 1$	33.9	774	784	$779 \pm 1$	52.8	
	10	636	613	$625 \pm 1$	10.4	1131	1143	$1137 \pm 1$	31.2	
Camphor	20	485	505	$495 \pm 1$	29	803	842	$822 \pm 1$	50.2	
	30	395	400	$397 \pm 1$	43.1	655	683	$669 \pm 1$	59.5	
	10	598	592	$595 \pm 1$	14.7	934	932	$933 \pm 1$	43.5	
Garlic	20	475	452	$463 \pm 1$	33.6	772	776	$774 \pm 1$	53.1	
	30	336	321	$328 \pm 1$	53	641	665	$653 \pm 1$	60.4	
	10	671	649	$660 \pm 1$	5.4	1188	1199	$1193 \pm 1$	27.8	
Mint	20	494	477	$485 \pm 1$	30.5	821	838	$830 \pm 1$	49.7	
	30	429	402	$416 \pm 1$	40.4	726	757	$741 \pm 1$	55.1	
Bellis® 38% V	WG	78	68	$73 \pm 1$	89.5	191	138	$165 \pm 1$	90	
Control		721	675	$698 \pm 1$	0.0	1652	1653	$1653 \pm 1$	0.0	
LSD at 0.05 %	)									
Treatment(T)		28.9	24.7			27	32.3			
Concentration	(C)	10.9	7.8			16.1	12.4			
(T x C)		25.1	20.5			42.5	32.8			

Efficacy of antioxidants to natural sugar beet powdery mildew: All tested resistance inducers caused significant reduction in AUPMPC values when sugar beet plants were sprayed with these tested inducers (Table 3). Increasing inducers concentration increased resistance of sugar beet plants against powdery mildew disease. The lowest AUPMPC on both cultivars Sirona and FD.0807 respectively was achieved by 300 ppm of salicylic acid followed by 200 ppm of salicylic acid and 300 ppm of citric acid respectively. The highest AUPMPC was obtained by 100 ppm of ascorbic acid. The best treatment (300 ppm salicylic acid) was higher in AUPMPC than the tested fungicide Bellis® 38% WG. The same trend was observed on the both growing seasons.

		AUPMPC								
Treatment	Conc. (ppm)		Sirona	cv.		FD.0807 L.				
		2014/2015	2015/2016	Mean	Protection (%)	2014/2015	2015/2016	Mean	Protection (%)	
	100	700	664	$682 \pm 1$	2.2	1100	1077	$1089 \pm 1$	34.1	
Ascorbic acid	200	648	638	$643 \pm 1$	7.8	1003	1013	$1008 \pm 1$	39	
	300	572	558	$565 \pm 1$	19	885	880	$882 \pm 1$	46.6	
	100	604	562	$583 \pm 1$	16.4	940	946	943 ±1	42.9	
Citric acid	200	521	503	$512 \pm 1$	26.6	782	798	$790 \pm 1$	52.2	
	300	470	471	$470 \pm 1$	32.6	707	718	$713 \pm 1$	56.8	
	100	537	516	$527 \pm 1$	24.4	813	820	$816 \pm 1$	50.6	
Salicylic acid	200	343	341	$342 \pm 1$	51	604	594	$599 \pm 1$	63.7	
	300	309	295	$302 \pm 1$	56.7	485	486	$485 \pm 1$	70.6	
Bellis® 38% W	G	78	68	$73 \pm 1$	89.5	191	138	$165 \pm 1$	90	
Control		721	675	$698 \pm 1$	0.0	1652	1653	$1653 \pm 1$	0.0	
LSD at 0.05 %										
Treatment(T)		40	25.5			32.5	31.1			
Concentration(C	C)	6.2	15.4			16.6	18.3			
(T x C)		14	34.4			37.2	41			

Table 3: Mean of AUPMPC values to sugar beet (Sirona& FD.0807 cultivars) as affected by foliar spray with antioxidants under field conditions during 2014/2015 and 2015/2016 growing seasons.

Efficacy of micronutrients to natural sugar beet powdery mildew: All tested microelements caused significant reduction in AUPMPC values when sugar beet plants were sprayed with any of these tested compounds (Table 4). Increasing microelements concentration increased resistance of sugar beet plants against powdery mildew disease. The lowest AUPMPC value on Sirona cultivar was achieved by 40 ppm of cupric sulfate followed by 20 ppm of cupric sulfate and 40 ppm of combination respectively. The highest AUPMPC value was obtained by 10 ppm of zinc sulfate. The best treatment (40 ppm, cupric sulfate) was higher in AUPMPC value than the tested fungicide Bellis® 38% WG. Data also showed that, the lowest AUPMPC value on FD.0807 cultivar was achieved by 40 ppm of cupric sulfate followed by 40 ppm of combination and 40 ppm of ferrous sulfate respectively. The highest AUPMPC value was obtained by 10 ppm of magnesium sulfate. The best treatment (40 ppm, cupric sulfate) was higher in AUPMPC value than the tested fungicide Bellis® 38% WG. The same trend was observed on the both growing seasons.

#### Discussion

The present study was carried out to evaluate ten sugar beet cultivars for their reaction to powdery mildew disease and to investigate the effect of five plant extracts *i.e.* basil, camphor, garlic, black cumin and mint at 10, 20, and 30 % concentrations on AUPMPC caused by Erysiphe betae. Also, the effect of three antioxidants (i.e. ascorbic acid, salicylic acid and citric acid) at 100, 200 and 300 ppm and four microelements (i.e. cupric sulfate. ferrous sulfate, magnesium sulfate, zinc sulfate and mixture of all of them) at the concentrations of 10, 20 and 30 ppm were studied Bellis® 38% WG fungicide applied was at the

#### recommended concentration to be used as a comparative treatment.

		AUPMPC							
Treatment	Conc. ppm	Sirona cv.				FD.0807 L.			
		2014/2015	2015/2016	Mean	Protection %	2014/2015	2015/2016	Mean	Protection %
	10	437	385	411 ±1	41.1	741	755	$748 \pm 1$	54.7
Cupric sulfate	20	217	209	$213 \pm 1$	69.4	417	436	$426 \pm 1$	74.2
	40	206	184	$195 \pm 1$	72	348	355	$352 \pm 1$	78.7
	10	487	499	$493 \pm 1$	29.3	831	847	$839 \pm 1$	49.2
Ferrous sulfate	20	294	309	$302 \pm 1$	56.7	523	514	$518 \pm 1$	68.6
	40	255	268	$261 \pm 1$	62.6	422	415	$418 \pm 1$	74.7
	10	556	518	$537 \pm 1$	23	934	928	931 ±1	43.6
Magnesium sulfate	20	390	356	$373 \pm 1$	46.5	673	676	$674 \pm 1$	59.2
-	40	346	314	$330 \pm 1$	52.7	593	572	$582 \pm 1$	64.7
	10	662	623	$642 \pm 1$	8	926	883	$904 \pm 1$	45.3
Zinc sulfate	20	431	414	$423 \pm 1$	39.3	727	696	$711 \pm 1$	56.9
	40	357	336	$347 \pm 1$	50.2	605	602	$603 \pm 1$	63.5
Combination	10	454	436	$445 \pm 1$	36.2	782	815	$799 \pm 1$	51.6
CS+FS+MS+ZS	20	263	250	$257 \pm 1$	63.1	487	467	477 ±1	71.1
C5   1 5   M5   25	40	246	225	$236 \pm 1$	66.1	414	377	$396 \pm 1$	76
Bellis® 38% WG		78	68	$73 \pm 1$	89.5	191	138	$165 \pm 1$	90
Control		721	675	$698 \pm 1$	0.0	1652	1653	$1653 \pm 1$	0.0
LSD at 0.05 %									
Treatment (T)		13.9	22.7			22.2	19.3		
Concentration (C)		12	11.3			15.8	17.3		
(T x C)		31.8	29.7			41.9	45.8		

Table 4: Mean of AUPMPC values to sugar beet (Sirona & FD.0807 cultivars) as affected by foliar spray with micronutrients under field conditions during 2014/2015 and 2015/2016 growing seasons.

The results indicate that Sirona (453 AUPMPC) was significantly the most tolerant cultivar to powdery mildew disease while, FD.0807 (1484 AUPMPC) was significantly the most susceptible cultivar, these results were in accordance with those reported by several researchers that concluded that plant cultivars are variants in their reactions to powdery mildew infection (Matsuda & Takamatsu, 2003; Abd-ElKareem et al., 2001). Fungicides have been used for a long time as the main strategy for controlling powdery mildew disease on sugar beet and subsequently increase vield production (Hassan & Berger, 1980; Docea & Fratila, 1980; Kontaxis, 1978 ; Abol-Wafa et al., 1976), Bellis® 38% WG fungicide when applied at the recommended concentration gave the best protection from powdery mildew on both sugar beet cultivars Sirona and FD.0807 respectively (89.5% & 90%). the highest effect of Bellis fungicide could be attributed to its mode of action due to its active ingredients which are related to the chemical fungicidal groups (Succinate dehydrogenase inhibitors and Quinone outside Inhibitors) those affect the respiration process in the fungal cell, those groups were found to be very effective against powdery mildew fungi in previous studies of (Karaoglanidis & Karadimos, 2006; Hollomon & Wheeler, 2002; Bartlett et al., 2002). On the other hand the fungicides resistant races of some pathogens have been reported by 2009; (Fernández-Aparicio et al.. Weiland & Koch, 2004). As well as the side effects of fungicides on human 82

health (Durmusoglu et al., 1997; Eckert & Ogawa, 1988) and the environment (Garcia, 1993; Horst et al., 1992). Despite that, fungicides are still the most dependable method in controlling such diseases although all their undesirable effects. Hence, there is a growing trend many years ago to involve other successful methods in disease management program depending on compounds which are useful, nontoxic and safe either on human health or on the environment. Among the tested plant extracts, the highest protection (69.9% & 66.9%) on both cultivars Sirona and FD.0807 respectively was achieved by 30% of basil extract followed by 30% of garlic extract (53% & 60.4%) while, the least protection (33.9% & 52.8%) was obtained by 10% of black cumin seed extract. The effect of plant derived constituents could be attributed to changes in plant metabolism which are dependent on the plant (Schmitt, 2002), such constituents lead to increase phenylalanine ammonia lyase activity in some plant leaves (Singh & Prithiviraj, 1997), ajoene (a compound derived from garlic) was used to control powdery mildew (Singh et al., 1995), the host plants exhibit delayed senescence and increases in chlorophyll contents. ethylene production and various enzyme activities were found in some treated plants (Herger & Klingauf, 1990). The importance of plant derived agents is not only for the inhibitory effect on the pathogen, but in way due to their ability to induce host resistance through increasing the activity of many enzymes which playing a defense role against invading pathogens (Nawar & Kuti, 2003; Caruso al., 2001). The et compounds responsible for the

preventative and curative effects could be fraction from these agents in relation to host resistance. Concerning the tested inducers, the highest protection (56.7% & 70.6%) was achieved by 300 ppm of salicylic acid followed by 200 ppm of salicylic acid (51% & 63.7%), while, 100 ppm of ascorbic acid gave the least protection (2.2% & 34.1%). Data are in agreement with those reported by several when they used researchers such compounds against several plant diseases caused by various pathogens (Sklodowska et al., 2010; Vimala et al., 2009; El-Samawaty & Galal, 2009; Ismail et al., 2006; Shaat & Galal, 2004; Sparla et al., 2004; Galal & Abdou, 1996). The reason for that effectiveness could be explained that many factors may act on plants to induce high levels of systemic resistance to subsequent pathogen attack. Some chemicals like benzothiadiazole, β-aminobutyric acid, chitosan, salicylic acid or even some plant extracts have been reported to induce resistance in a number of plantpathogen interactions that can be very effective (Barilli et al., 2009,2010; Bélanger & Labbé, 2002; Dann & Deverall, 2000; Frey & Carver, 1998). They may provide commercially useful broad-spectrum plant protection that is stable, long-lasting and environmentally benign. Exogenous application of salicylic acid solutions to pea leaves induced systemic resistance to E. pisi and the percentages of fungal germlings that successfully infected untreated leaves were reduced by 20-30% (Frey & Carver, 1998). On the other hand, 40 ppm of cupric sulfate achieved the best percentage of protection (72% & 78.7%) whereas, the least protection (8%) was obtained by 10 ppm of zinc sulfate on Sirona cultivar and the least protection (43.6%) was obtained by 10 ppm of magnesium sulfate on FD.0807 cultivar. Finally, it is recommended to conduct further studies on the possibilities of applying useful compounds such as plant derived compounds, resistance inducers, macro and micro nutrients in the open field and to produce them commercially in order to reduce the risk of fungicides on human health and environment.

## References

- Abd-El-Kareem F, Abd\_Alla MA, El-Mohamedy RSR, 2001. Induced resistance in potato plants for controlling late blight disease under field conditions. Egyptian Journal of Phytopathology **29**: 29–41.
- Abol-Wafa MT, Farag SA, Kamara AM, Shehata MR 1976. Fungicidal control of cucumber powdery mildew. Egyptian Journal of Phytopathology **8**: 47–54.
- Barilli E, Sillero JC, Rubiales D, 2009. Induction of systemicacquired resistance in pea against rust (*Uromycespisi*) byexogenous application of biotic and abiotic inducers. Journal of Phytopathology **158**: 30–34.
- Barilli E, Prats E, Rubiales D, 2010. Benzothiadiazole and BABA improve resistance to Uromycespisi (Pers.) wint.in Pisumsativum L. with an enhancement of enzymatic activities and total phenolic content. European Journal of Plant Pathology **128**: 483–493.
- Bartlett DW, Clough JM, Godwin JR, Hall, AA, Hamer M, Parr-Dobrzanski B, 2002: The strobilurin fungicides. Pest Management Science **58**: 649–662.

- Bélanger RR, Labbé C, 2002. Control of powdery mildews without chemicals: prophylactic and biological alternatives for horticulturalcrops. In: Bélanger, R.R.; Bushnell, W.R.; Dik, A.J. and Carver, T.L.W. (eds.) The powdery mildews, a comprehensive treatise. APS Press, St. Paul, MN, USA, 256-267 pp.
- Caruso C, Chilosi G, Leonard L, Bertin L, Magro P, Buonocore V, Caporal C, 2001. A basic peroxidase from wheat kernel with antifungal activity. Phytochemistry 58: 743–750.
- Chiha F, Morjane H, Harrabi M, 1997. Use of area under the disease progress curve to screen potential parents for *Ascochyta* blight resistance. In: Udupa SM, Weigand F (eds.). DNA Markers and Breeding for Resistance to *Ascochyta* Blight in Chickpea. Proceeding of the Symposium on 'Application of DNA Fingerprinting for Crop Improvement: Marker-Assisted Selection of Chickpea for Sustainable Agriculture in Dry Areas', Aleppo 1994, 209-215.
- Dann EK, Deverall BJ, 2000. Activation of systemic diseaseresistance in pea by an avirulent bacterium or a benzothiadiazole, but not by a fungal leaf spot pathogen. Plant Pathology 49:324–332.
- Docea E, Fratila E, 1980. A new pathogen causing powdery mildew of cucumber in Romania. Lucrari Stiintifice, B. **22**: 165– 168.
- Durmusoglu E, Massfeld W, Sengonca C, 1997. Determination of the exposure of workers to two different pesticides in a greenhouse with roses. Mitteilungen der Deutschen Gesellschaft fur Allgemeine und Angewandte Entomologie **11**(1/6): 319–322.

- Eckert JW, Ogawa JM, 1988. The chemical control of postharvest diseases, deciduous fruits, berries, vegetable and root/tuber crops. Annual Review of Phytopathology **26**: 433–469.
- Elad Y, 1992. The use of antioxidants (free radical scavengers) to control grey mould (*Botrytis cinerea*) and white mould (*Sclerotiniasclerotiorum*) in various crops. Plant Pathology **41**: 417–426.
- El-Fahhar Samia A, 2008. Impact of powdery mildew disease on root weight, sugar yield and purity of sugar beet. Journal Agricultural Research - Kafr El-Sheikh University **34**(3): 636–646.
- El-Fahar Samia A, Mostafa Sahar ME, 2009: Effect of some plant extracts and partial substitution of thiovit 80 by plant extract on powdery mildew disease of sugar beet. Journal of Agricultural Science -Mansoura University 34(1): 309–320.
- El-Samawaty AMA, Galal AA, 2009. Use of benthiodiazole (BTH) for inducing systemic resistance in cotton seedlings against some soil-borne pathogenic fungi. Journal of Agricultural Science -Mansoura University **34**(4): 3305–3315.
- Fernández-Aparicio M, Prats E, Emeran AA, Rubiales D, 2009. Characterization of resistance mechanisms to powdery mildew (*Erysiphebetae*) in beet (*Beta vulgaris*). Phytopathology **99**: 385–389.
- Francis SA, 2002. Sugar-beet powdery mildew (*Erysiphebetae*). Molecular Plant Pathology **3**:119–124.
- Frey S, Carver TLW, 1998. Induction of systemic resistance in pea to pea powdery mildew by exogenous application of salicylic acid. Journal of Phytopathology **146**: 239–245.

- Gado EAM, 2013. Impact of treatment with some plant extracts and fungicides on sugar beet powdery mildew and yield components. Australian Journal of Basic and Applied Sciences **7**(1): 468–472.
- Galal AA, Abdou ElS, 1996. Antioxidants for the control of fusarial disease in cowpea. Egyptian Journal of Phytopathology **24**: 1–12.
- Garcia JE, 1993. Pesticides as contaminants. Turrialba (Costa Rica), **43**(3): 221–229.
- Gomez KA, Gomez AA, 1984: Statistical procedures for agricultural research, Second edition, John Wiley & Sons, Inc., New York, USA, 680 pp.
- Hassan SA, Berger H, 1980: Integrated control on glasshouse vegetable crops. Institut für biologische Schädlingsbekämpfung, D- 6100 Darmstadt, German Federal Republic, Workshop sessions, 485–486 pp.
- Hassan TI, 2006. The biological control via biofertilizers. M.Sc. Thesis, Faculty of Agriculture, Minia University, Minia, Egypt, 135 pp.
- Herger G, Klingauf F, 1990. Control of powdery mildew fungi with extracts of the giant knotweed, *Reynoutriasachalinensis*, Polygonaceae. Mededelingen van de Faculteit Landbouwwetenschappen, Rijksuniversiteit Gent **55**(3a): 1007–1014.
- Hills FJ, Chiarappa L, Geng S, 1980. Powdery mildew of sugar beet: Disease and crop lossassessment. Phytopathology **70**: 680–682.
- Hogenboom NG, 1993. Economic importance of breeding for disease resistance, Indurability of disease resistance, Kluwer Dordrecht, 5–9 pp.

- Hollomon DW, Wheeler IE, 2002. Controlling powdery mildews with chemistry. In: Bélanger RR, Bushnell WR, Dik AJ, Carver T.W (eds.), The powdery mildews, a comprehensive treatise. APS Press, St. Paul, MN, USA, 249–255 pp.
- Horst RK, Kawamoto SO, Porter LL, 1992. Effect of sodium bicarbonate and oils on control of powdery mildew and black spot of roses. Plant Disease **76**: 247–251.
- Ismail ME, Abdalla HM, Galal AA, 2006. Factors affecting induced resistance in sunflower plants against basal stem rot caused by *Sclerotium rolfsii* (*Corticumrolfsii*). Minia Journal of Agricultural Research and Development 26(3): 405–425.
- Ismail ME, Galal AA, Abdalla HM, 2012: Induction of resistance in sunflower plants against powdery mildew. Minia international conference for agriculture and irrigation in the Nile basin countries  $26^{\text{th}} - 29^{\text{th}}$  March 2012, Minia, Egypt.
- Karaoglanidis GS, Karadimos DA, 2006. Efficacy of strobilurinsand mixtures with DMI fungicides in controlling powdery mildew infield-grown sugar beet. Crop Protection **25**: 977–983.
- Kontaxis Demetrios G, 1978: Bayleton® and sulfur control sugar beet powdery mildew in Imperial Valley California, disease effect and yield. Phytopathology News **12**: 143–144.
- Kontradowitz L, Verreet JA, 2010. Assessment of resistance and virulence in the pathosystem sugar beet (*Beta vulgaris*) / powdery mildew (*Erysiphebetae*) – development of basics for an effective powdery mildew resistancebreeding Journal of Plant Diseases and Protection **117** (2): 49–54.

- Kuc J, 1995. Phytoalexins, stress metabolism, and disease resistance in plants. Annual Review of Phytopathology 33: 275–297.
- Matsuda S, Takamatsu S, 2003. Evolution of host-parasite relationships of *Golovinomyces* (Ascomycete: Erysiphaceae) inferred from nuclear rDNA sequences. Molecular Phylogenetics and Evolution **27**: 314– 327.
- Moharam MHA, Obiadalla Ali HAE, 2012. Preventative and curative effects of several plant derived agents against powdery mildew disease of okra. Notulae Scientia Biologicae 4(3): 76–82.
- Moustafa MSH, Abdel-Shahid YA, Ez-Eldin I, Anwar HM, 1990. Factors affecting acquired resistance in *Erysiphe cichoracearum*, the causal organism of cucumber powdery mildew. Agricultural Research Review **68** (3):521–528.
- Nawar HF, Kuti JD, 2003. Wyerone acid phytoalexin synthesis and peroxidase activity as markers for resistance of broad beans to chocolate spot disease. Journal of Phytopathology **151**:564–570.
- Radwan KSA, 2017. Integrated management of tomato wilt and root rot diseases, PhD thesis, Faculty of Agriculture, Minia University, Minia, Egypt, 140 pp.
- Schmitt A, 2002. Induced responses by plant extracts from *Reynoutria sachalinensis*, a case study, Bulletin OILB/SROP **25**(6): 83–88.
- Shaat MNM, Galal AA, 2004. Response of citrus fruits to pre-harvest to antioxidants spraying and infection with *Alternaria* fruit rot and green mold. Annals of Agricultural Sciences **49**: 747–758.

- Singh UP, Prithiviraj B, 1997. Neemazal, a product of neem (*Azadirachta indica*) induces resistance in pea (*Pisum sativum*) against *Erysiphe pisi*. Physiological and Molecular Plant Pathology **51**:181–194.
- Singh UP, Prithiviraj B, Wagner KG, Plank-Schumacher K, 1995. Effect of ajoene, a constituent of garlic (*Allium sativum*), on powdery mildew (*Erysiphepisi*) of pea (*Pisum sativum*). Journal of Plant Diseases and Protection **102**(4): 399– 406.
- Sklodowska M, Gajewska E, Kuzniak E, Mikicinski A, Sobiczewski P, 2010. BTH-mediated antioxidant system responses in apple leaf tissues. Scientia Horticulture **125**: 34–40.
- Sparla F, Rotino L, Valgimigli MC, Pupillo P, Trost P, 2004. Systemic resistance induced by benzothiadiazole in pear inoculated with the agent of fire blight (*Erwinia amylovora*). Scientia Horticulture **101**(3): 269–279.

- Vimala R, Suriachandraselvan M, 2009. Induced resistance in bhendi against powdery mildew by foliar application of Salicylic acid, Journal of Biopesticides 2(1): 111–114.
- Weiland JJ, Koch G, 2004. Sugar-beet leaf spot disease (*Cercospora beticola* Sacc.). Molecular Plant Pathology **5**: 157–166.
- Whitney ED, Lewellen RT, Skoyen IO, 1983. Reaction of sugar beet to powdery mildew: Genetic variation, association among testing procedures, and results of resistance breeding. Phytopathology **73**: 182–185.